

## Expression of activation antigens CD69, HLA-DR, interleukin-2 receptor- $\alpha$ (IL-2R $\alpha$ ) and IL-2R $\beta$ on T cells of human decidua at an early stage of pregnancy

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### SUMMARY

T cells of human decidua at an early stage of pregnancy were examined by flow cytometry for the expression of the T-cell-activation antigens CD69, HLA-DR, interleukin-2 receptor- $\alpha$  (IL-2R $\alpha$ ) and IL-2R $\beta$ . The decidua contained a small number of T cells and both CD4<sup>+</sup> and CD8<sup>+</sup> subsets expressed CD69, HLA-DR, IL-2R $\alpha$  and IL-2R $\beta$  antigens significantly whereas, in peripheral blood, only a small number of T cells expressed these activation antigens. These findings indicate that T cells in the decidua in the first trimester of pregnancy are regionally activated.

Since the decidualized endometrium is the site of fertilized ovum implantation and placental invasion, mechanisms for the maintenance of pregnancy may be elucidated by examining decidual tissue for immunocompetent cells.

Recent studies have suggested that maternal T cells play an important role in maintaining pregnancy in the mouse.<sup>1–6</sup> Wegmann proposed that immunostimulation, rather than suppression, is important in maintaining pregnancy. This concept, termed the 'immunotrophism hypothesis', is based on the report that a soluble T-lymphocyte factor [i.e. granulocyte-macrophage colony-stimulating factor (GM-CSF) and interleukin-3 (IL-3)] enhances the proliferation of murine trophoblast cells *in vitro*. If the immunotrophism model is valid in humans, T cells in the decidua should be in an activated state, because these cytokines are not secreted from T cells in the resting state. Therefore, we examined whether human intradecidual T cells express the activation antigens, CD69, HLA-DR, interleukin-2 receptor- $\alpha$  (IL-2R $\alpha$ ) and IL-2R $\beta$  by flow cytometric analysis.

Tissue samples were obtained from 12 women who had indicated elective abortions of normal pregnancy between the sixth and twelfth week of pregnancy under informed consent. Decidual tissue and peripheral blood samples were taken from each woman when the abortions were performed. The decidual tissue was macroscopically separated from chorionic villi, and then cut into small pieces and vigorously shaken for 1 min without enzymatic treatment. These samples were then filtered through a 32- $\mu$  nylon mesh, and decidual mononuclear cells were isolated by the Ficoll-Hypaque gradient sedimentation.

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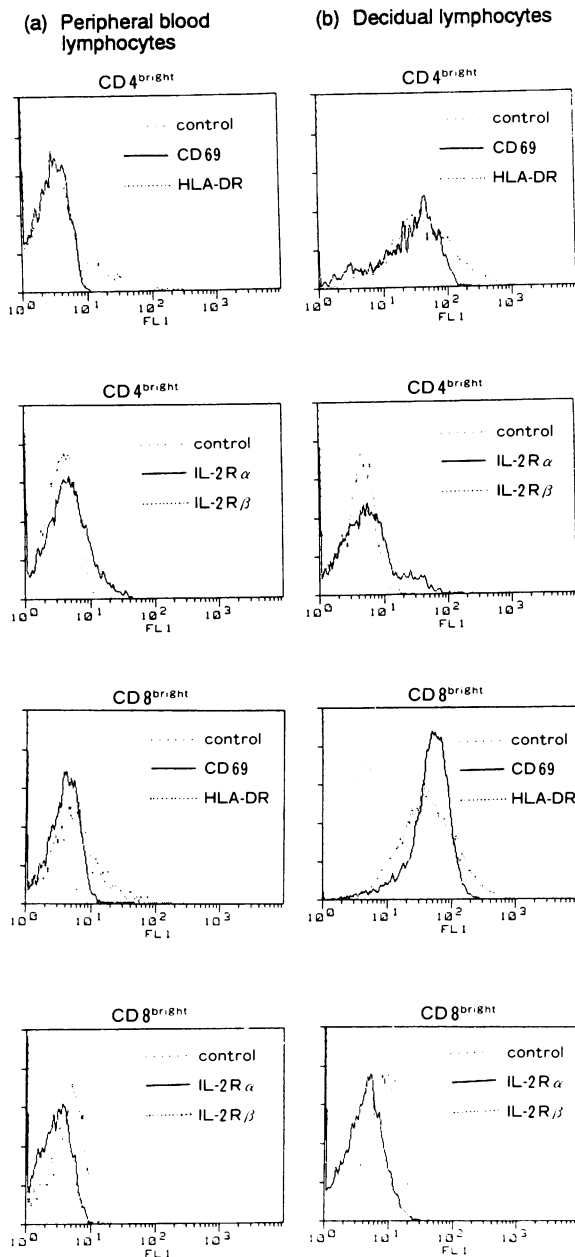
Peripheral blood mononuclear cells (PBMC) were also isolated by the standard Ficoll-Hypaque method.

The following monoclonal antibodies (mAb), purchased from Becton Dickinson Immunocytometry System (San Jose, CA), were used: (1) CD3 (Leu-4), CD4 (Leu-3), CD8 (Leu-2), CD14 (Leu-M3), CD16 (Leu-11), CD20 (Leu-16) and CD56 (Leu-19); and (2) CD69 (Leu-23), HLA-DR (HLA-DR), CD25; IL-2R $\alpha$ , p55(2A3) and CD45 (HLe-1). The first seven of these mAb were labelled with phycoerythrin (PE), and the last four and TU27 (IL-2R $\beta$ , p75) mAb, which was kindly given by Dr Sugamura (Tohoku University, Japan), were labelled with fluorescein isothiocyanate (FITC).

The size of decidual lymphocytes ranged from that of resting peripheral blood lymphocytes to monocytes, and thus decidual macrophages were unable to be excluded with scatter gate. The lymphocyte population, identified by forward and size scatter gates, contained 86.8  $\pm$  6.6% (mean  $\pm$  SD,  $n$  = 12) lymphocytes and 6.6  $\pm$  4.1% (mean  $\pm$  SD,  $n$  = 12) macrophages, as analysed by two-colour flow cytometry with CD45-FITC and CD14-PE.

Flow cytometry was performed on a FACScan (Becton Dickinson). The number of cells counted was 50,000. Positive rates were expressed as percentages of positive cells in all lymphocytes (excluding monocytes/macrophages).

The percentage of CD16<sup>+</sup> CD56<sup>bright</sup> natural killer (NK) cells, which have been called endometrial granulocytes,<sup>7–10</sup> was 84.8  $\pm$  5.2% of the decidual lymphocytes, whereas the percentage of CD16<sup>+</sup> NK cells was very low (2.7  $\pm$  1.2%). The percentage of CD3<sup>+</sup> T cells was 12.2  $\pm$  4.6%, which was markedly lower than that in peripheral blood. These cell populations are similar to those described in previous studies.<sup>8–10</sup> The percentages of CD4<sup>bright</sup> cells and CD8<sup>bright</sup> cells were 5.6  $\pm$  2.1% and 4.7  $\pm$  1.9%, respectively. The CD4/CD8 ratio was 1.23  $\pm$  0.42.



**Figure 1.** Expression of CD69, HLA-DR, IL-2R $\alpha$  and IL-2R $\beta$  on peripheral blood CD4<sup>bright</sup> cells, peripheral blood CD8<sup>bright</sup> cells, decidua CD4<sup>bright</sup> cells and decidua CD8<sup>bright</sup> cells. PBL and decidua lymphocytes were stained with FITC-conjugated anti-CD69, HLA-DR, IL-2R $\alpha$  and IL-2R $\beta$  and PE-conjugated anti-CD4 or CD8. Samples were analysed by FACScan and displayed as histograms. Since decidua macrophages were unable to be excluded with the scatter gate, we set the marker tightly on each subset, CD4<sup>bright</sup> and CD8<sup>bright</sup> cells.

The expression of the CD69 antigen (an early activation marker of T and NK cells)<sup>11,12</sup> by decidua CD4<sup>bright</sup> and CD8<sup>bright</sup> cells was  $58.3 \pm 12.7\%$  and  $73.3 \pm 14.3\%$ , respectively. On the other hand, peripheral blood CD4<sup>+</sup> and CD8<sup>+</sup> cells in early pregnant women express CD69 in only  $0.9 \pm 0.5\%$  and  $3.5 \pm 0.7\%$  of cells, respectively (Fig. 1, Table 1). CD69 antigen has been reported to appear shortly after stimulation via protein kinase C (PKC). Our finding indicates that the T cells in the

**Table 1.** T-cell activation marker expression on decidua and peripheral blood T cells in the first trimester of pregnancy

	Decidual T cells (n = 12)	Peripheral blood T cells (n = 12)
CD4:CD69	$58.3 \pm 12.7^{****}$	$0.9 \pm 0.5$
HLA-DR	$67.7 \pm 14.8^{****}$	$11.4 \pm 4.6$
IL-2R $\alpha$	$26.9 \pm 5.0^*$	$20.5 \pm 5.2$
IL-2R $\beta$	$8.6 \pm 4.6^{***}$	$1.5 \pm 0.5$
CD8:CD69	$73.3 \pm 14.3^{****}$	$3.5 \pm 0.7$
HLA-DR	$82.9 \pm 13.5^{****}$	$18.6 \pm 8.0$
IL-2R $\alpha$	$6.2 \pm 2.9^{**}$	$2.5 \pm 1.5$
IL-2R $\beta$	$17.7 \pm 7.2^*$	$10.6 \pm 3.4$

† % of CD4<sup>+</sup> or CD8<sup>+</sup> cells. All values are mean  $\pm$  SD.

Statistical significances were noted between decidua T cells and peripheral blood T cells: \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ ; \*\*\*\* $P < 0.0001$ .

decidua of pregnant women have already been stimulated via PKC.

HLA-DR antigen (a relatively late activation T-cell marker) was detected in  $11.4 \pm 4.6\%$  of CD4<sup>+</sup> cells and in  $18.6 \pm 8.0\%$  of CD8<sup>+</sup> cells in peripheral blood lymphocytes of pregnant women (Table 1). In contrast, in the decidua, most T cells showed HLA-DR antigen expression ( $67.7 \pm 14.8\%$  of CD4<sup>+</sup> cells and  $82.9 \pm 13.5\%$  of CD8<sup>+</sup> cells) (Table 1, Fig. 1). Thus, we show here that decidua T cells express an early activation antigen, CD69, and a late activation antigen, HLA-DR. In relation to this, it is interesting to know to what extent IL-2 receptors are expressed. It has been reported that a proportion of resting CD4<sup>+</sup> T cells express IL-2R $\alpha$  and resting CD8<sup>+</sup> T cells express IL-2R $\beta$  slightly.<sup>13,14</sup> However, decidua CD4<sup>+</sup> cells express not only IL-2R $\alpha$  but also IL-2R $\beta$ . Decidua CD8<sup>+</sup> T cells express not only IL-2R $\beta$  but also IL-2R $\alpha$ . The level of antigen expression was slight but significant (Fig. 1, Table 1). These results were not caused by contamination with either CD56<sup>bright</sup> cells, which possess IL-2R $\alpha$  and IL-2R $\beta$ ,<sup>10-15</sup> or CD16<sup>+</sup> cells and monocytes which possess IL-2R $\beta$ ,<sup>13,14,16</sup> because we set the marker tightly on each subset, CD4<sup>bright</sup> and CD8<sup>bright</sup> cells. Bulmer and Johnson reported that, using an immunohistochemical method, decidua lymphocytes did not possess IL-2R $\alpha$ .<sup>17</sup> In our recent flow cytometric study, we observed the expression of both IL-2R $\alpha$  and IL-2R $\beta$  by CD16<sup>-</sup> CD56<sup>bright</sup> NK cells, the major population of decidua lymphocytes.<sup>10</sup> The discrepancy between our study and other earlier studies is probably attributable to the higher sensitivity of flow cytometry compared with immunohistochemical methods.

In this study, both CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the decidua expressed the activation antigens CD69, HLA-DR, IL-2R $\alpha$  and IL-2R $\beta$ . It has been reported that anti-CD8 treatment of pregnant mice reduces placental proliferation and phagocytosis, and that anti-CD4 antibody treatment affects placental proliferation.<sup>5</sup> Thus, it would appear that activated CD4- and CD8-positive cells may be involved in placental cell proliferation and function.

Another important question concerns the mechanism of T-cell activation in the decidua. Since trophoblast produces an IL-2-like molecule and expresses IL-2 messenger RNA (mRNA) at

an early stage of pregnancy,<sup>18,19</sup> it is possible that this factor may activate T cells. However, it is also possible that other antigen-presenting cells in the decidua are involved in T-cell activation.<sup>20</sup> Immunosuppressive factors, such as transforming growth factor-beta (TGF- $\beta$ ) and prostaglandin E<sub>2</sub>, are known to be distributed regionally in the decidua.<sup>21,22</sup> These factors could influence the expression of IL-2R $\alpha$  and IL-2R $\beta$ , because expression of these activation markers was slight compared with CD69 and HLA-DR. In the present study, T cells appeared phenotypically to be in an activated state, even in the presence of immunosuppressive factors. Pregnancy thus seems to be maintained by the regulation of immunosuppressive and immunopotentiating mechanisms.

The next stage of investigation should address the question of whether activated T cells in the human decidua secrete cytokines (e.g., IL-3 and GM-CSF) in the area of fertilized egg implantation.

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